The Synthesis and Pharmacological Activity of 5-Ethyl-5-(3-hydroxy-1-methylbutyl)barbituric Acid¹

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The anesthetic, pentobarbital, is excreted chiefly as 5-ethyl-5-(3-hydroxy-1-methylbutyl)barbituric acid^{2a} (III). Studies using pentobarbital-2-C¹⁴ show that over 70% of the drug is hydroxylated.^{2b} It has been stated that these metabolic products have no pharmacological activity.³ With the synthesis (Scheme I) of adequate quantities of material, the pharmacological properties of III have been reinvestigated.



Pharmacological Data.—Female, Swiss–Webster mice were treated with the test compounds suspended in 1% Methocel and administered orally by stomach tube. Two hours after drug administration, the animals were tested by the maximal electroshock method of Toman, Swinyard, and Goodman.⁴ Diphenylhydantoin at a dose of 10 mg/kg was found to be more effective than III at a dose of 500 mg/kg in protecting against the tonic extensor component of convulsions.

By intraperitoneal administration of III as a 1% Methocel suspension, the ED₅₈ in the maximal electroshock test was found to be 310 (252–381) mg/kg. No indication of anesthesia or ataxia was noted even in doses of 1 g/kg.

It is concluded from these experiments that III has a very weak anticonvulsant activity and no anesthetic properties. It is doubtful if any of the action

(3) R. T. Williams, "Detoxification Mechanisms," John Wiley and Sons, Inc., New York, N. Y., 1959, p 600.

(4) J. E. P. Toman, E. A. Swinyard, and L. S. Goodman, J. Neurophysiol., 9, 231 (1946). of pentobarbital can be ascribed to accumulations of the metabolite.

Experimental Section

Diethyl Ethyl(1-methyl-3-oxobutyl)malonate (I).—Diethyl ethylmalonate (188 g, 1.0 mole) was added to a stirred solution of sodium (1.3 g, 0.056 g-atom) in 150 ml of dry ethanol at 25° . 3-Penten-2-one (67.2 g, 0.8 mole), prepared by the method of Alexander and Coraor,⁶ was added at 10° over a 1-hr period. The mixture was stirred at 10° for 2.5 hr, then neutralized with acetic acid. It was added to 300 ml of water, and the oil layer was separated. The aqueous layer was extracted with two 50-ml portions of ether. The ether extract and the oil layer were combined, and the solvent was removed by distillation. The residue was distilled to yield I (193 g, 88.7%), bp 102-103° (0.5 mm), n^{35} D 1.4440. The product assayed 98% by glc. Infrared and nmr spectra were consistent with the assigned structure.

Anal. Calcd for $C_{14}H_{24}O_{5}$: C, 61.8; H, 8.85. Found: C, 61.7; H, 8.91.

Ethyl Ethyl(3-hydroxy-1-methylbutyl)malonate δ -Lactone (II). —A solution of I (29.5 g, 0.11 mole) in 100 ml of absolute ethanol was reduced with Raney Ni at 3 atm of H₂. The solution was filtered and the solvent removed by distillation. Distillation of the residue gave II (20 g, 79%), bp 102° (0.1 mm), $n^{c_{5}}$ D 1.4560. Infrared and nmr spectra were consistent with the assigned structure. Purity was 98% by glc.

Anal. Calcd for $C_{12}H_{20}O_4$: C, 63.2; H, 8.77. Found: C, 62.9; H, 8.81.

5-Ethyl-5-(3-hydroxy-1-methylbutyl)barbituric Acid (III).— Urea (15 g, 0.25 mole) was added to a stirred solution of sodium (5.75 g, 0.25 g-atom) in 150 ml of dry ethanol. This was stirred to solution at 40°, and then II (19 g, 0.09 mole) was added over a 30-min period. The solution was then refluxed for 40 hr. The solvent was removed at 20 mm until the pot temperature reached 50°. The residue was dissolved in 200 ml of water at 5–10° and extracted with two 50-ml portions of ether. The aqueous solution was neutralized to a pH of 6.0 with 5 N HCl and the dissolved ether was removed under reduced pressure. The solution was cooled and filtered to give III (10.9 g, 51.2%), mp 170–175°. The crude product was recrystallized from water to give 6.7 g of III, mp 187–188° (uncor). Infrared and mmr spectra were consistent with the assigned structure.

Anal. Calcd for $C_{11}H_{16}N_2O_3$: C, 54.5; H, 7.45; N, 11.58. Found: C, 54.7; H, 7.43; N, 11.26.

Samples of III were compared with material extracted from the urine of a cat anesthetized with pentobarbital. The urine was acidified to pH 6.4 and extracted with ethyl acetate. Samples were chromatographed on Whatman No. 1 paper with 1-butanol saturated with 1% ammonia as the descending solvent. The chromatograms were sprayed with a 0.1% saturated solution of cobalt acetate in pyridine. The barbiturate derivatives gave a dull purple color with this reagent. The $R_{\rm f}$ of the synthetic material in this system was 0.80 which compares favorably with the material ($R_{\rm f}$ 0.78) extracted from the urine.

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(5) E. R. Alexander and G. R. Coraor, J. Am. Chem. Soc., 73, 2721 (1951).

The Synthesis of 3-Fluoroestra-1,3,5(10)-trienes

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The substituent at the 3-position of the steroidal estrogens plays an important role in the pharmacological activities of these compounds. A vast improvement in the ratio of the hypocholesterolemic and geno-

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(a) E. W. Maynert and H. B. van Dyke, J. Pharmacol. Exptl. Therap., 98, 184 (1950); E. W. Maynert and J. M. Dawson, J. Biol. Chem., 195, 389 (1952); (b) E. Titus and H. Weiss, *ibid.*, 214, 807 (1955).

Notes

tropic¹ activities to the estrogenic activity has been achieved by the elimination of the 3-hydroxyl group of the natural estrogens² or by the preparation of various aminoalkyl ethers of this group,³ respectively. These changes have greatly lowered the estrogenicity of these compounds, which still retain a relatively large proportion of their antiovulatory and/or hypocholesterolemic activities. The introduction of fluorine atoms onto the nucleus of a variety of steroid hormones had led, in several instances, to additional or greatly enhanced activities for these compounds.⁴ The substitution of a fluorine atom for the oxygen function at the 3position of the estratriene nucleus should lead to an interesting comparison of the effects of these two small, highly electronegative atoms.

3-Fluoroestra-1,3,5(10)-trien-17-one (III) was prepared by a Schiemann reaction of 3-animoestra-1,3,5-(10)-trien-17-one (II).⁵ Other 3-substituted estratrienes have previously been prepared by Hecker⁶ by replacement of the corresponding diazonium ion prepared from 17β -acetoxy-3-aminoestra-1,3,5(10)triene. The starting aniline II was readily prepared from estrone in 58% over-all yield by the general method developed by Scherrer for the conversion of phenols into anilines.⁷ This synthesis seems to be superior, from the standpoint of both yield and convenience, to that previously employed for the preparation of this compound.⁵ Estrone was condensed with 4-chloro-2-phenylquinazoline⁸ to give 3-(2-phenyl-4-quinazolinyloxy)estra-1,3,5(10)-trien-17-one (IV). which was pyrolyzed at 330° to yield 3-(4-oxo-2phenyl-3(4H)-quinazolinyl)estra-1,3,5(10)-trien-17-oue (V). This material was hydrolyzed to give 3-aminoestra-1,3.5(10)-trien-17-one (II), which was converted



(1) M. L. Black, J. Pharm. Pharmacol., 13, 127 (1961).

A. H. Goldkamp, W. H. Hoelm, R. A. Mikulec, E. F. Nutting, and D. L. Cook, J. Med. Chem., 8, 409 (1965).

(3) D. D. Evans, D. E. Evans, G. S. Lewis, P. J. Palmer, and D. J. Weyell, J. Pharm. Pharmacol., 16, 717 (1964).

(4) N. Applezweig, "Steroid Drugs," McGeaw-Hill Book Co., Inc., New York, N. Y., 1962.

(5) A. M. Gold and E. Schwenk, J. Am. Chem. Soc., 81, 2108 (1959).

(6) E. Hecker, Chem. Ber., 95, 977 (1962).

(7) R. A. Scherrer, Abstracts of Papers, 145th National Meeting of the Meeting Chemical Society, New York, N. Y., Sept 1963, p 33Q.

(8) M. M. Endicott, E. Wick, M. L. Mercury, and M. L. Sherrill, J. Am. Chem. Soc., 68, 1239 (1946). to III by a standard Schiemann reaction.⁹ A similar process was previously used to convert 1-hydroxy-4methylestra-1,3,5(10)-trien-17-one to the corresponding 1-fluoro derivative.¹⁰ Sodium borohydride reduction of III yielded the estradiol derivative VI, and treatment of III with methylmagnesinm bromide afforded the corresponding 17α -methyl analog VII.

These three 3-fluoroestra-1,3,5(10)-tricnes (III, VI, and VII) as well as the aromatic amine II were inactive as genotropic agents¹ when assayed orally in mice at 5 mg/kg/day.¹¹ No oral hypocholesterolemic activity was found for these four compounds in rats at 2.5 mg/kg/day.¹² The estradiol analog VI had no dc-tectable antiestrogenic activity and was a very weak estrogen when compared with estradiol in rats (*ca.* 1/100).¹³

Experimental Section¹⁴

3-Aminoestra-1,3,5(10)-trien-17-one (II).—A solution of 26.4 μ (0.098 mole) of estrone in 1 1, of dried diglyme was treated with 4.89 g (0.104 mole) of a 51% dispersion of NaH in mineral oil. The mixture was stirred nuder an atmosphere of nitrogen until the evolution of hydrogen ceased. To this solution was then added 25.3 g (0.105 mole) of 4-chloro-2-phenylquinazolite,⁸ and the resulting mixture was stirred and refluxed under nitrogen for 1.5 hr. The cooled mixture was poured into ice-water, and the product was collected by filtration, washed well with water, and dried under reduced pressure. The crude **3-(2-phenyl-4-quin-azolinyloxy)estra-1,3,5(10)-trien-17-one** (IV), which weighed 50.45 g and exhibited λ_{max} 256 m μ (ϵ 31,000) and 286 m μ (ϵ 16,200), was used directly for the next step.

A suspension of 25.2 g of crude IV in 250 ml of heavy minerai oil was covered with an atmosphere of nitrogen, stirred, and heated at 330–335° for 5 hr. The solution was then cooled to room temperature, diluted with 500 ml of petroleum ether tbp 35-60°, and filtered. The crude **3**-(**4**-oxo-2-phenyl-3(4H)-quinazolinyl)estra-1,3,5(10)-trien-17-one (V), which weighed 25.0 g and exhibited λ_{max} 279 m μ (ϵ 14,100), was used directly for the next step.

A solution of 25.0 g of crude V in 1500 ml of absolute ethanol was treated with 350 ml of 40% aqueous NaOH solution and refluxed for 7 hr. This solution was then cooled in ice, treated with 650 ml of 12 N HCl, and allowed to stand overnight at room temperature. The mixture then was stirred and refluxed for 1.5 hr, cooled, and filtered. The NaCl precipitate was washed well with ethanol and discarded. The filtrate and washings were concentrated under reduced pressure, poured into water, and filtered. The filtrate was made alkaline with NaOH solution, saturated with K₂CO₅, and filtered. The precipitate was washed well with water, dried under reduced pressure, and sublimed at 165° (0.1 mm). The sublimate was recrystallized from benzenecyclohexane to yield 7.65 g of 11, mp 193–195° (lit.⁵ mp 192– 192.6°). The yield of product based upon starting estrone was 58%.

3-Fluoroestra-1,3,5(10)-trien-17-one (III). —A solution of 1,00 g (3.72 mmoles) of 3-aminoestra-1,3,5(10)-trien-17-one (11) in 20 ml of ethanol and 10 ml of 48% fluoroboric acid was cooled to 0°, treated with a solution of 0.26 g (3.77 mmoles) of NaNO₂ in 1 ml of water, and stirred at 0–5° under an atmosphere of nitrogen for 1.5 hr. The solution was then diluted with 1 l. of ether and filtered. The precipitated diazoninm fluoroborate saft was dried

- (10) D. F. Morrow and M. E. Butler, J. Ocg. Chem., 29, 1893 (1964).
- (11) R. Q. Thompson, M. Stortevani, and O. D. Bird, Science, 118, 657 (1953).
- (12) G. Rodney, M. L. Black, and O. D. Bird, Biochem. Phychonesist, 14, 445 (1965).
- (13) The methods used are essentially those of G. D. Duncan, S. C. Lyster, J. J. Clark, and D. Ledniver [*Proc. Soc. Exptl. Biol. Med.*, **112**, 439 (1063)] with a slight molification in the antiestorgen assay in which the test morpound and estradiol were administered for 7 days beginning 8 days after ovarieetomy.

(14) Melting points were determined on a Fisher-Johns block and are corrected. The altraviolet spectra were run in mericand. Optical rotations were determined on a 1% solution in CHCl₅. All compounds had infrared spectra which agreed with their assigned structures.

⁽⁹⁾ A. Roe, Org. Reactivits, 5, 193 (1949).

under reduced pressure at 70°, at which temperature it decomposed to give the crude 3-fluoro compound. This crude material was dissolved in benzene, filtered, and chromatographed on alumina (Woelm, neutral, activity grade I). The product was eluted with 10% ether in benzene and recrystallized from methanol to give 0.24 g (24%) of III, mp 178–180°, $[\alpha]^{24}$ D +143°. Anal. Calcd for C₁₈H₂₁FO: C, 79.38; H, 7.77; F, 6.98.

Found: C, 79.39; H, 8.00; F, 6.83.

3-Fluoroestra-1,3,5(10)-trien-17β-ol (VI).—A solution of 1.00 g (3.69 numbers) of 3-fluoroestra-1,3,5(10)-trien-17-one (III) in 100 ml of ethanol was treated with 4 ml of 10% NaOH solution and 0.53 g (14.0 mmoles) of NaBH₄. After 1 hr at room temperature the solution was poured into water, and the product was extracted with ether. The extract was washed with water, dried, and concentrated to dryness. The residue was recrystallized from hexane to yield 0.65 g (65%) of VI, mp 112-114°, [a]²⁴D +84°.

Anal. Caled for C18H23FO: C, 78.80; H, 8.45; F, 6.92. Found: C, 79.00; H, 8.71; F, 6.67.

3-Fluoro-17 α -methylestra-1,3,5(10)-trien-17 β -ol (VII).—A solution of 0.84 g (3.09 mmoles) of 3-fluoroestra-1,3,5(10)-trien-17one (III) in 80 ml of ether was treated with 3.5 ml of 3M nethyl-magnesium bromide solution. The resulting mixture was stirred and refluxed for 1 hr, cooled, and treated with NH4Cl solution. The ether layer was separated, washed with water, dried, and concentrated to dryness. The residue was recrystallized from ether-petroleum ether to give 0.40 g (45%) of VII, mp 108-110°, $[\alpha]^{24}D + 58^{\circ}$.

Anal. Calcd for C₁₉H₂₅FO: C, 79.13; H, 8.74; F, 6.59. Found: C, 78.90; H, 8.75; F, 6.24.

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Synthesis of Some Substituted Tryptophols of Possible Physiological Importance and a Study with 3-(2-Acetoxyethyl)-5-methoxyindole (5-Methoxytryptophol O-Acetate) on Sexual Maturation

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The unique psychopharmacological effects of ethanol, easily formed by biological processes, have been appreciated since antiquity. Recent studies have indicated that hydroxyl derivatives of histanine,^{2a} the catecholamines,^{2b,e} γ -aminobutyric acid,³ tryptamine,⁴ and serotonin⁵ can be metabolites of these biogenic anines. Furthermore, the resultant biogenic alcohols may possess physiological properties. An example is γ -hydroxybutyric acid, a metabolite of γ -aminobutyric acid³ which has effective sleep-producing properties.⁶

Tryptophol was identified as a metabolite of tryptanine in rats pretreated by the aldehyde dehydrogenase blocking agent, disulfiram.⁴ Kveder, et al.,⁵ presented good evidence that 5-hydroxytryptophol may be the second most important metabolite of serotonin in the rat, rather than N-acetylserotonin as first thought.⁷ Bartholini, Pletscher, and Bruderer⁸ found a neutral metabolite of serotonin released from isolated blood platelets by reserpine and have presented chromatographic evidence that it is 5-hydroxytryptophol. Evidence has also been presented for the presence of 5-methoxytryptophol and 5-hydroxytryptophol in pineal tissue⁹ rather than β -carbolines, once thought to be constituents.¹⁰

It was necessary for us to prepare a collection of substituted tryptophols, the acetate esters, and related acids for our studies of metabolism, isolation, and pharmacology of these substances. Acetylation alters markedly the hormonal properties of 5-methoxytryptanine. The latter is without effect on frogskin lightening while the acetylated analog (melatonin) is a potent frog-skin-lightening hormone^{11,12} and displays marked inhibitory effects on the incidence of estrus in rats.¹³ Therefore, it was of importance to see if chain acetylation of 5-methoxytryptophol might also produce a compound with special biological properties not seen in the precursor.

5-Methoxytryptophol was prepared by the lithium aluminum hydride reduction of either 5-methoxyindole-3-glyoxyloyl chloride or 5-methoxyindole-3-acetic acid. The reduction of the former produced a considerable amount of by-product which could not be removed by distillation or fractional crystallization. However, the 5-methoxytryptophol could be purified by conversion to a solid picrate derivative. Reduction of the acid gave a better product which was easily purified without recourse to the picrate. It was a yellow oil which was acetylated with acetic anhydride to give an oil that did not solidify, but that could be converted into a useful picrate. The metabolic fate and effects on sexual development of 3-(2-acetoxyethyl)-5-methoxyindole in the female rat were investigated; the latter study is reported in this paper, and the former will be reported elsewhere.

Other substituted tryptophols prepared were conpounds which could be potential metabolites of 5methoxytryptophol O-acetate and serve as chroniatographic standards. For the selection of authentic compounds to be synthesized for chromatography, three possible metabolic transformations were anticipated. These are described together with synthetic routes (Chart I) being: (1) 6-hydroxylation only to give VII; (2) 6-hydroxylation and hydrolysis to give V; (3) 6-hydroxylation, ester hydrolysis, and oxidation to give III. Usually, indole-3-acetonitriles can

⁽¹⁾ Research Fellow supported by Training Grant 5126-08 from the National Institutes of Health, U. S. Public Health Service.

^{(2) (}a) T. Nakajima and I. Sane, Biochim. Biophys. Acta, 82, 260 (1964); (b) J. Axelrod, I. J. Kopin, and J. P. Mann, *ibid.*, **36**, 576 (1959); (c) M. Goldstein, A. J. Freidhoff, S. Pomerantz, and C. Simmons, ibid., 39, 189 (1960).

⁽³⁾ S. P. Bessman and W. N. Fishbein, Nature, 200, 1207 (1963).

⁽⁴⁾ A. A. Smith and S. B. Wortis, Biochim. Biophys. Acta, 40, 569 (1960).

⁽⁵⁾ S. Kveder, S. Iskric, and D. Keglevic, Biochem. J., 85, 447 (1962).

⁽⁶⁾ H. Laborit, J. M. Jounay, J. Gerard, and F. Fabiani, Presse Med., 68, 1867 (1960); M. Blumenfeld, R. Suntay, and M. Harmel, Anesthesia Analgesia, Current Res., 44, 721 (1962).

⁽⁷⁾ W. M. McIsaac and I. H. Page, J. Biol. Chem., 234, 858 (1959).

⁽⁸⁾ G. Bartholini, A. Pletscher, and H. Bruderer, Nature, 203, 1281 (1964).

⁽⁹⁾ W. M. McIsaac, G. Farrell, R. G. Taborsky, and A. N. Taylor, Science, 148, 102 (1965).

⁽¹⁰⁾ G. Farrell and W. M. McIsaac, Arch. Biochem. Biophys., 94, 543 (1961).

⁽¹¹⁾ A. B. Lerner and J. D. Case, J. Invest. Dermatol., 32, 211 (1959). (12) A. B. Lerner, J. D. Case, and R. V. Heinzelman, J. Am. Chem. Soc., 81, 6084 (1959).

⁽¹³⁾ R. J. Wurtman, J. Axelrod, and E. W. Chu. Science, 141, 277 (1963).